

# Buffers and Ionic Salts: Densities and Solubilities of Aqueous and Electrolyte Solutions of Tris(hydroxymethyl)aminomethane and *N*-Tris[hydroxymethyl]-4-amino-butanesulfonic Acid<sup>†</sup>

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The densities of aqueous solutions of tris(hydroxymethyl)aminomethane (TRIS) and tris[hydroxymethyl]-4-amino-butanesulfonic acid (TABS), useful biological buffers within the pH range of 7.0 to 9.0 for TRIS and 8.2 to 9.5 for TABS, have been measured by a high-precision vibrating-tube digital densitometer in aqueous and in aqueous electrolyte solutions from (298.15 to 328.15) K under atmospheric pressure. This study was undertaken to investigate the interactions between these compounds and electrolytes of potassium acetate (KAc), potassium bromide (KBr), potassium chloride (KCl), and sodium chloride (NaCl). In this series of measurements, the aqueous samples were prepared with various concentrations of the buffers, up to saturated conditions, and over salt concentrations from (1 to 4) mol·dm<sup>-3</sup>. The experimental densities were correlated as a function of the concentration of the buffers and ionic salts. The solubilities of buffers at 298.15 K in aqueous and in aqueous electrolyte solutions have also been determined from the experimental results of density measurements. It was found that the solubilities of TRIS and TABS in aqueous solution decrease with increasing concentration of salts (salting-out effect). The solubility data were further used to estimate the apparent free energy of transfer ( $\Delta G_{tr}'$ ) of buffers from water to aqueous electrolyte solutions. The contribution of TABS residue ( $-CH_2CH_2CH_2CH_2SO_3^-$ ) from water to aqueous electrolyte solutions was predicted from the  $\Delta G_{tr}'$  results. The measured densities served to evaluate the apparent molar volumes,  $V_{\phi}(m, T)$ , and fitted them to an equation that describes the surface ( $V_{\phi}$  against  $T$  against  $m$ ). The apparent molar volumes of buffers at infinite dilution ( $V_{\phi}^{\circ}$ ) have been determined from the solubility data. The trends of transfer volumes ( $\Delta_{tr}V_{\phi}^{\circ}$ ) have been interpreted in terms of solute–cosolute interactions on the basis of a cosphere overlap model.

## Introduction

Buffers are ubiquitous components in most in vitro reaction systems because many biological and chemical systems involve acid–base equilibria and therefore depend critically on the pH of the solution. Buffers covering the pH range between 6 and 9 are especially required for biological and environmental studies. Many early buffers were not suitable for biological applications because they may not be inert enough, thus interfering with the system under study. The recent extension of the number of buffers covering the physiological pH range now makes it possible to assess much more effectively the suitability of possible buffer systems.

*N*-Tris[hydroxymethyl]-4-amino-butanesulfonic acid (TABS) is a zwitterionic buffer which gives good activity of four enzymes with optimum pH values in the alkaline range:  $\beta$ -galactosidase, esterase, phosphodiesterase, and alkaline phosphatase.<sup>1</sup> In biological systems where effects of K<sup>+</sup> and Na<sup>+</sup> may be important, buffers lacking these ions may be desirable; such buffers may comprise amines including tris(hydroxymethyl)aminomethane (TRIS)<sup>2</sup> and its derivatives and the zwitterionic “Good” buffers provided by Good and his associates.<sup>3,4</sup> TRIS-buffer is extensively used in biochemistry and molecular biology.<sup>5</sup> In biochemistry, TRIS is widely used

as a component of buffer solutions, such as in TRIS-borate-EDTA (TBE) and TRIS-acetate-EDTA (TAE) buffers, especially for solutions of nucleic acids. TRIS salts are used in protein crystallization at various pH values.<sup>6–9</sup> Currently, most researchers use TRIS-buffer as an extender for canine semen preservation.<sup>10,11</sup> Canine semen can be the most efficiently frozen with an extender constituted by TRIS-buffer.<sup>12</sup> TRIS has been utilized in studies of double stranded complexes of peptide nucleic acids (PNA) and their cDNA sequences.<sup>13</sup>

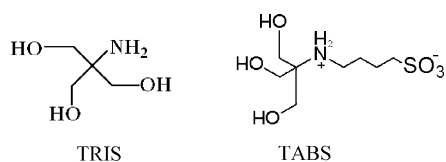
TRIS buffers have been widely used to define seawater's pH values. Early workers<sup>14</sup> used TRIS buffers in natural seawater as secondary pH standards. TRIS buffers in synthetic seawater are now used as primary buffers to define seawater pH scales<sup>15–18</sup> either directly for subsequent calibration of electrometric pH measurements<sup>19,20</sup> or indirectly by using them to define the p*K* of indicator dyes for seawater pH measurements.<sup>21–23</sup>

Although TRIS has been a major biochemical buffer for many years, partly because it is relatively inexpensive and readily available in a highly purified form, it has disadvantages. These include its reactivity as a primary amine and its appreciable solubility in organic solvents which leads to its accumulation in the biological phases of reaction systems. Thus, TRIS buffer displaces the electron transport, and phosphorylation, pH unit when compared with a number of other buffers. It also inhibits isocitrate dehydrogenase of pea mitochondria, whereas 4-(2-hydroxyethyl)piperazine-1-ethanesulfonic acid (HEPES) does not. The TBE and TAE buffers form complexes with DNA.<sup>24–27</sup>

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## Scheme 1



DNA molecules also form extensive complexes with histidine molecules in isoelectric histidine buffers.<sup>28</sup> The complexes can be dissociated by adding monovalent salts such as NaCl or KBr to the solution, indicating that the complexes are stabilized by electrostatic interactions. Divalent cations are less effective than monovalent cations in dissociating the complexes. DNA–TRIS interactions also influence the rate of cleavage of plasmid pBR322 by the restriction enzyme EcoRV.<sup>29</sup>

With the aim to contribute to the knowledge of the interactions of these buffers (Scheme 1) with electrolytes, the present work reports the interactions between four selected salts (KAc, KBr, KCl, and NaCl) and two biological buffers of TRIS and TABS, via density measurements, which were measured by a high-precision vibrating-tube digital densitometer. We have determined the solubility of TRIS and TABS in water and in aqueous electrolyte solutions at 298.15 K from the results of the density measurement. In addition, these data can be used to study the apparent free energy of transfer of these buffers from water to aqueous electrolyte solutions at 298.15 K. We calculated the apparent molar volumes,  $V_{\phi}(m, T)$ , from the solubility data.

### Experimental Section

**Materials.** TRIS (mass fraction purity > 0.999) and TABS (mass fraction purity > 0.99) were purchased from Sigma Chemical Co. (USA). The salts, potassium chloride (KCl, mass fraction purity 0.9999+) and potassium acetate (KAc, mass fraction purity 0.9998+), were purchased from Aldrich Chemical Co. (USA). Potassium bromide (KBr, mass fraction purity 0.995+) and sodium chloride (NaCl, mass fraction purity 0.995+) were obtained from Arcos Organics (USA). All the purchased materials were used without further purification. Water used for making the aqueous and aqueous electrolyte solutions was obtained from the NANO pure-Ultra pure water system that was distilled and deionized with resistance of 18.3 M $\Omega$ . All the aqueous solution samples were prepared gravimetrically.

**Density Measurements.** Densities were measured with an Anton Paar DMA-4500 vibrating-tube densitometer, Austria, with an uncertainty of  $\pm 5 \cdot 10^{-5} \text{ g} \cdot \text{cm}^{-3}$ . For this kind of instrument, the sample fills a vibrating tube. Usually, it is assumed that the densitometer behaves as an oscillator without damping. This is true for low-viscosity samples, but for high-viscosity ones the damping contribution is significant, a fact that makes the “measured” density higher than the real one.<sup>30</sup> Therefore, corrections for high-viscosity samples are needed for this kind of densitometer. There are some studies that deal with this problem, in which different procedures are proposed to make such corrections.<sup>31</sup> The main advantage of the DMA-4500 densitometer is its ability to carry out this correction automatically, without any prior information about the viscosity of the sample. The densitometer is equipped with an internal temperature control unit, which can regulate the temperature of the measuring tube to within  $\pm 0.03 \text{ K}$  in a temperature range of (273.15 to 363.15) K. The instrument was calibrated with air and degassed distilled water.

**Solubility Measurements.** Solubilities of TRIS and TABS were measured at 298.15 K from the density measurements,

following the procedure of Nozaki and Tanford.<sup>32–34</sup> The detailed procedure used in this work has been delineated in our earlier articles.<sup>35,36</sup> Briefly, at least nine sample vials were prepared for each investigated system. To each of the sample vials containing a fixed amount of solvent (water or aqueous electrolyte solutions) was added weighed amounts of a buffer compound to provide a series of mixtures with increasing composition of buffer mass. Of the nine vials, four to six contained low enough weights of buffer such that unsaturated solutions resulted. The remaining vials contained sufficient buffer weights to produce saturated solutions, and each of the vials was then sealed with a Teflon-coated screw cap. The vials were completely in a thermostatic shaker equipped with a water bath (BT-350R, Yih-Der, Taiwan) at  $T = 298.15 \text{ K}$  for (36 to 48) h, and the supernatant of saturated solutions was removed through a syringe and filtered by a 0.22  $\mu\text{m}$  disposal filter (Millipore, Millex-GS) before performing the density measurements. In the present study, (0, 1, 2, and 4)  $\text{mol} \cdot \text{dm}^{-3}$  of KAc, KBr, KCl, or NaCl aqueous solutions were chosen as solvents. Since the densities of liquid samples will remain constant as the amounts of added buffers are greater than their solubility limit, the saturated solubility can be determined by the composition where density levels off on a plot of density against the original amount of buffer added in the solvent. The uncertainty of the solubility limit is lower than  $\pm 0.8 \%$ . The concentration of buffer in the samples, in units of (g of buffer/100 g of solvent), is calculated from eq 1.

$$\text{composition } \frac{\text{g}_{\text{buffer}}}{100 \text{ g}_{\text{solvent}}} = \left( \frac{\text{weight}_{\text{buffer}}(\text{g})}{\text{weight}_{\text{solvent}}(\text{g})} \right) \cdot 100 \quad (1)$$

The solubility limits expressed as (g of buffer/100 g of solvent) are converted to the more appropriate units of molarity (moles of buffer/liter of solution) and molality (moles of buffers/kg of solvent) using eqs 2 and 3, respectively.<sup>37</sup>

$$\text{molality} = \frac{W_{\text{B}} 1000}{M_{\text{B}} W_{\text{sv}}} \quad (2)$$

$$\text{molarity} = \frac{W_{\text{B}} \rho^*}{M_{\text{B}} (W_{\text{B}} + W_{\text{sv}})} \quad (3)$$

where  $W_{\text{B}}$  is the weight of buffer in grams;  $W_{\text{sv}} = 100 \text{ g}$  of solvent;  $M_{\text{B}}$  is the buffer molecular weight; and  $\rho^*$  is the density ( $\text{g} \cdot \text{L}^{-1}$ ) of the saturated solution at the solubility limit determined experimentally. To check the reliability of the experimental procedure used in the present study, we measured the solubility limits ( $S_{\text{B}}$ ) of TRIS buffer in water at  $T = 298.15 \text{ K}$  ( $S = 5.76 \text{ mol} \cdot \text{kg}^{-1}$ ), which agree fairly well with the values  $5.780 \text{ mol} \cdot \text{kg}^{-1}$  and  $5.766 \text{ mol} \cdot \text{kg}^{-1}$  reported by Bates et al.<sup>38</sup> and El-Harakany et al.,<sup>39</sup> respectively.

### Results and Discussion

The measured experimental densities ( $\rho$ ) for the aqueous electrolyte solutions at several temperatures are shown in a previous paper.<sup>40</sup> It was found that the densities of the electrolyte solutions increase with increasing unity of molarity of salt and decrease with temperature, and the salt effects on density increment follow the order of  $\text{KBr} > \text{KAc} \approx \text{KCl} > \text{NaCl}$ . Tables 1 and 2 present the densities and concentrations of buffers in





Table 3. Predicted Results from Equation 4 and Correlated Results from Equation 6 for Ternary Aqueous Systems

system	$10^2 \cdot a_1$	$a_2$	$p_1^b$	$p_2^b$	$p_3^b$	from eq 1	from eq 3
	$\text{g} \cdot \text{cm}^{-3a}$	$\text{g} \cdot \text{cm}^{-3a}$				AARD % <sup>c</sup>	AARD % <sup>d</sup>
water + TRIS	3.15602	0.708699	—	—	—	0.11	—
water + TABS	7.74654	0.661610	—	—	—	0.26	—
water + TRIS + KAc <sup>e</sup>	3.15602	0.708699	-0.38966	0.525332	0.743455	1.04	0.16
water + TRIS + KBr <sup>e</sup>	3.15602	0.708699	-1.34417	0.571821	1.052408	2.00	0.14
water + TRIS + KCl <sup>e</sup>	3.15602	0.708699	-0.12753	0.318879	0.543195	1.14	0.20
water + TRIS + NaCl <sup>e</sup>	3.15602	0.708699	-0.41331	0.524147	0.863622	1.25	0.16
water + TABS + KAc <sup>e</sup>	7.74654	0.661610	-2.22187	0.775811	1.045978	2.05	0.27
water + TABS + KBr <sup>e</sup>	7.74654	0.661610	-3.13290	0.668427	1.185392	3.74	0.18
water + TABS + KCl <sup>e</sup>	7.74654	0.661610	-1.80870	0.730897	1.180139	1.98	0.15
water + TABS + NaCl <sup>e</sup>	7.74654	0.661610	-1.17204	0.673487	1.002374	2.08	0.21

<sup>a</sup> Parameters in eq 4. <sup>b</sup> Parameters in eq 6. <sup>c</sup> AARD/% =  $(100/N) \sum_{i=1}^N |\rho_i^{\text{calc}} - \rho_i^{\text{expt}}|/\rho_i^{\text{expt}}$ , where  $N$  is the number of data points and  $\rho^{\text{calc}} = \rho_w + a_1[m]^{a_2} + b_1[\text{salt}]^{b_2}$ . <sup>d</sup> AARD/% =  $(100/N) \sum_{i=1}^N |\rho_i^{\text{calc}} - \rho_i^{\text{expt}}|/\rho_i^{\text{expt}}$ , where  $N$  is the number of data points and  $\rho = \rho_w + a_1[m]^{a_2} + b_1[\text{salt}]^{b_2} + p_1[m/100]^{p_2}[\text{salt}/10]^{p_3}$ . <sup>e</sup> Parameters  $10^2 \cdot b_1$  and  $b_2$  are 4.62317 and 0.946424 for the KAc-containing system; 8.21748 and 0.984256 for the KBr-containing system; 4.40501 and 0.970168 for the KCl-containing system; and 3.92698 and 0.960618 for the NaCl-containing system, respectively.<sup>40</sup>

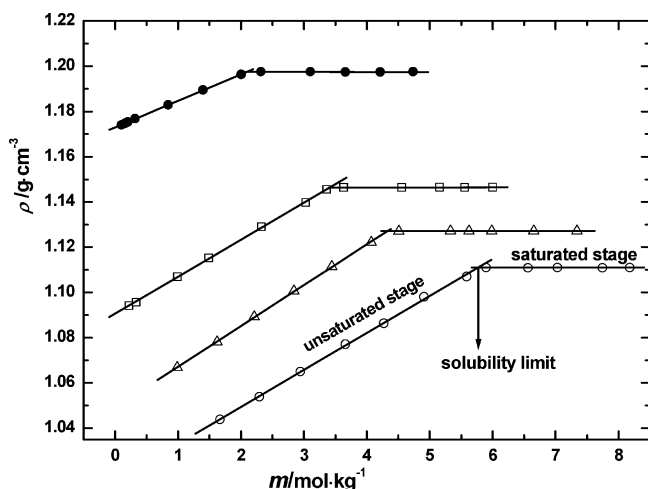


Figure 1. Representative solubility profiles of TRIS in water or KAc aqueous solutions at 298.15 K:  $\circ$ , water;  $\Delta$ ,  $1 \text{ mol} \cdot \text{dm}^{-3}$  KAc;  $\square$ ,  $2 \text{ mol} \cdot \text{dm}^{-3}$  KAc;  $\bullet$ ,  $4 \text{ mol} \cdot \text{dm}^{-3}$  KAc. The symbol  $m$  is the composition of buffer in the prepared sample vials.

For the salt-free ( $0 \text{ mol} \cdot \text{dm}^{-3}$ ) and the buffer-free (aqueous electrolytes) binary systems, the densities of solutions over the entire temperature range can be expressed satisfactorily by the following empirical equation<sup>40</sup>

$$\rho = \rho_w + a_1[m]^{a_2} + b_1[\text{salt}]^{b_2} \quad (4)$$

where  $\rho_w$  is the density of pure water;  $[m]$  is the concentration of buffers ( $\text{mol} \cdot \text{kg}^{-1}$ ); and  $[\text{salt}]$  is salt concentration in  $[\text{mol} \cdot \text{dm}^{-3}]$ . The optimal values of parameters  $a_1$  and  $a_2$  are reported in Table 3 for buffer-free binary systems, while  $b_1$  and  $b_2$  were previously reported.<sup>40</sup> The values of  $(a_1 \times a_2)$  in eq 4 represent the density increment caused by increasing a mole of the buffer in 1 kg of solvent. It shows that the magnitude of density increment of TABS is greater than that of TRIS. The tabulated average absolute deviation (AARD) is defined as (Table 3)

$$\text{AARD}/\% = (100/N) \sum_{i=1}^N |\rho_i^{\text{calc}} - \rho_i^{\text{expt}}|/\rho_i^{\text{expt}} \quad (5)$$

where  $N$  is the number of data points.

It was notable that the densities of the (water + buffers + salts) ternary systems could not be estimated directly from eq 4 with the determined parameters ( $a_1, a_2$ ) and ( $b_1, b_2$ ). The AARDs

of prediction from eq 4, as presented in Table 3, are obviously greater than the experimental uncertainty. The accuracy of density calculations was significantly improved by adding an extra term into eq 4 to take the interactions between buffer and salt into account; i.e.

$$\rho = \rho_w + a_1[m]^{a_2} + b_1[\text{salt}]^{b_2} + p_1 \left[ \frac{m}{100} \right]^{p_2} \left[ \frac{\text{salt}}{10} \right]^{p_3} \quad (6)$$

where  $p_1, p_2$ , and  $p_3$  are model parameters whose optimal values can be obtained by fitting the above empirical model to density data for each ternary system of (water + buffer + salt). The optimized values of  $p_1, p_2$ , and  $p_3$  together with the AARD of density correlation for each ternary system are presented in Table 3. This empirical model can correlate the density data to within about the experimental uncertainty. Similar calculated results were observed for the other temperatures as well. These results provide evidence for the existence of interactions between the buffers and ionic salts. As seen from Table 3, the values of parameter  $p_1$  are negative for all the investigated ternary systems, indicating that the interactions between buffers and ionic salts result in density decrease or volume expansion. It was found that the values of parameters  $p_1, p_2$ , and  $p_3$  of TABS are significantly greater than those on TRIS. This indicates that the zwitterionic TABS molecules are larger in size in solutions with higher salt concentrations. This effect can be attributed to the fact that the amino and sulfonic groups of TABS dissociate in aqueous solutions and become positively and negatively charged, respectively. The dissociation of TABS molecules in the presence of salts leads to the formation of ion pairs between the charged groups of TABS and salt ions. The formation of the ion pairs, on the one hand, increases the apparent molar volume of TABS and, on the other hand, reduces the electrostatic interactions between TABS molecules and water molecules and ions. This in turn reduces the interactions of water molecules and ions with the hydrocarbon backbone of TABS resulting in an increase in the number of water molecules attached to the amino and sulfonic groups of TABS in the form of hydration water. This effect can also be seen from the increasing values of  $V_{\phi}^{\circ}$  of TABS at higher salt concentrations (hereafter, Table 9).

The solubility of TRIS and TABS at  $T = 298.15 \text{ K}$  in water and in aqueous electrolyte solutions was determined from the results of density measurements. As an example, Figure 1 shows the density profile of TRIS in water and in (1, 2, and 4)  $\text{mol} \cdot \text{dm}^{-3}$  concentrations of KAc. The solubility limits, ex-

**Table 4. Buffer Solubilities ( $S_B$ ) and Densities ( $\rho^*$ ) at Solubility Limits in Water or Ionic Salt Aqueous Solutions at  $T = 298.15$  K and at Atmospheric Pressure**

solvent	$S_B/\text{mol}\cdot\text{kg}^{-1}$				$\rho^*/\text{g}\cdot\text{cm}^{-3}$			
	$0\text{ mol}\cdot\text{dm}^{-3}$	$1\text{ mol}\cdot\text{dm}^{-3}$	$2\text{ mol}\cdot\text{dm}^{-3}$	$4\text{ mol}\cdot\text{dm}^{-3}$	$0\text{ mol}\cdot\text{dm}^{-3}$	$1\text{ mol}\cdot\text{dm}^{-3}$	$2\text{ mol}\cdot\text{dm}^{-3}$	$4\text{ mol}\cdot\text{dm}^{-3}$
TRIS								
water	5.76 <sup>a</sup>				1.11099			
KAc		4.35	3.40	2.11		1.12716	1.14640	1.19750
KBr		4.76	3.92	2.46		1.15906	1.20929	1.31998
KCl		4.92	4.29	2.73		1.13123	1.15505	1.21403
NaCl		4.97	4.56	3.52		1.13123	1.15078	1.19350
TABS								
water	5.91				1.25825			
KAc		5.51	4.77	4.13		1.26499	1.28340	1.29957
KBr		5.54	5.21	4.50		1.29211	1.32772	1.39925
KCl		5.75	5.32	4.95		1.27639	1.29147	1.32565
NaCl		5.61	5.47	4.89		1.26982	1.28526	1.31216

<sup>a</sup>  $S_B = 5.780$ ;<sup>38</sup>  $5.766$ .<sup>39</sup>

**Table 5. Apparent Transfer Free Energies of TRIS, TABS, and TABS Residue Contribution from Water to Aqueous Electrolyte Solutions at  $T = 298.15$  K and at Atmospheric Pressure**

solvent	$\Delta G'_{tr}/\text{J}\cdot\text{mol}^{-1}$				$\Delta g'_{tr}/\text{J}\cdot\text{mol}^{-1}$		
	$1\text{ mol}\cdot\text{dm}^{-3}$	$2\text{ mol}\cdot\text{dm}^{-3}$	$4\text{ mol}\cdot\text{dm}^{-3}$	$1\text{ mol}\cdot\text{dm}^{-3}$	$2\text{ mol}\cdot\text{dm}^{-3}$	$4\text{ mol}\cdot\text{dm}^{-3}$	TABS residue ( $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{SO}_3^-$ )
TRIS							
KAc	396.42	770.20	1559.62	57.18			
KBr	183.23	396.19	1019.29	-0.81			
KCl	192.56	359.79	1026.99	-8.70			
NaCl	176.25	270.09	612.44	28.83			
TABS							
KAc					176.28	312.30	-339.24
KBr					-4.01	26.39	-184.04
KCl					41.06	54.95	-201.26
NaCl					24.83	92.06	-147.42
TABS residue ( $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{SO}_3^-$ )							
							-593.92
							-400.20
							-318.73
							-245.26
							-1247.32
							-992.90
							-972.04
							-520.38

pressed as  $S_B$  ( $\text{mol}\cdot\text{kg}^{-1}$ ), and the densities ( $\rho^*$ ) of saturated solution at solubility limit were obtained at the intersection of the two fitted lines in each experiment. Table 4 summarizes the solubility limits and the corresponding densities ( $\rho^*$ ) of the saturated solutions. The experimental results in Table 4 demonstrate that the solubilities of TRIS and TABS decrease with increasing concentrations of KAc, KBr, KCl, and NaCl solutions and indicate the salting-out effect is dominant. Among the series of potassium salts, the observed order of the salting-out effect is  $\text{KAc} > \text{KBr} > \text{KCl}$ . The anions exert their effects independently, which are the relative effects of  $\text{CH}_3\text{COO}^-$ ,  $\text{Br}^-$ , and  $\text{Cl}^-$ . Furthermore, we found that the salting-out effect of  $\text{KCl} > \text{NaCl}$  for the TRIS buffer and  $\text{NaCl} \approx \text{KCl}$  for TABS buffer, respectively.

The free energy of transfer of a solute from one solvent system to another can be based on the solubilities of that solute in these two solvent systems.<sup>32,41</sup> At the solubility limit of the biological buffer in aqueous electrolyte solutions, two phases are in equilibrium, the solid phase of the buffer and the liquid phase of solution. At equilibrium, the chemical potential of the biological buffer in the crystalline phase must be equal to the chemical potential of the biological buffer in solution. Since the chemical potential of a buffer in its crystal is invariant, regardless of the solvent with which it is in contact, and since this chemical potential is the same in both solvent systems, the chemical potential of the biological buffer at the saturation point in one solvent must equal its chemical potential at the saturation point in the second solvent,<sup>32-34,37,43,44</sup> eq 7.

$$\mu_{B,w}^o + RT \ln S_{B,w} + RT \ln \gamma_{B,w} = \mu_{B,ws}^o + RT \ln S_{B,ws} + RT \ln \gamma_{B,ws} \quad (7)$$

where  $\mu_{B,w}^o$  and  $\mu_{B,ws}^o$  are the standard chemical potentials assigned to the buffer in water and in salt solution;  $S_{B,w}$  and  $S_{B,ws}$  are the buffer solubilities in water and in salt solution and expressed either in molar, molal, or mole fraction units;

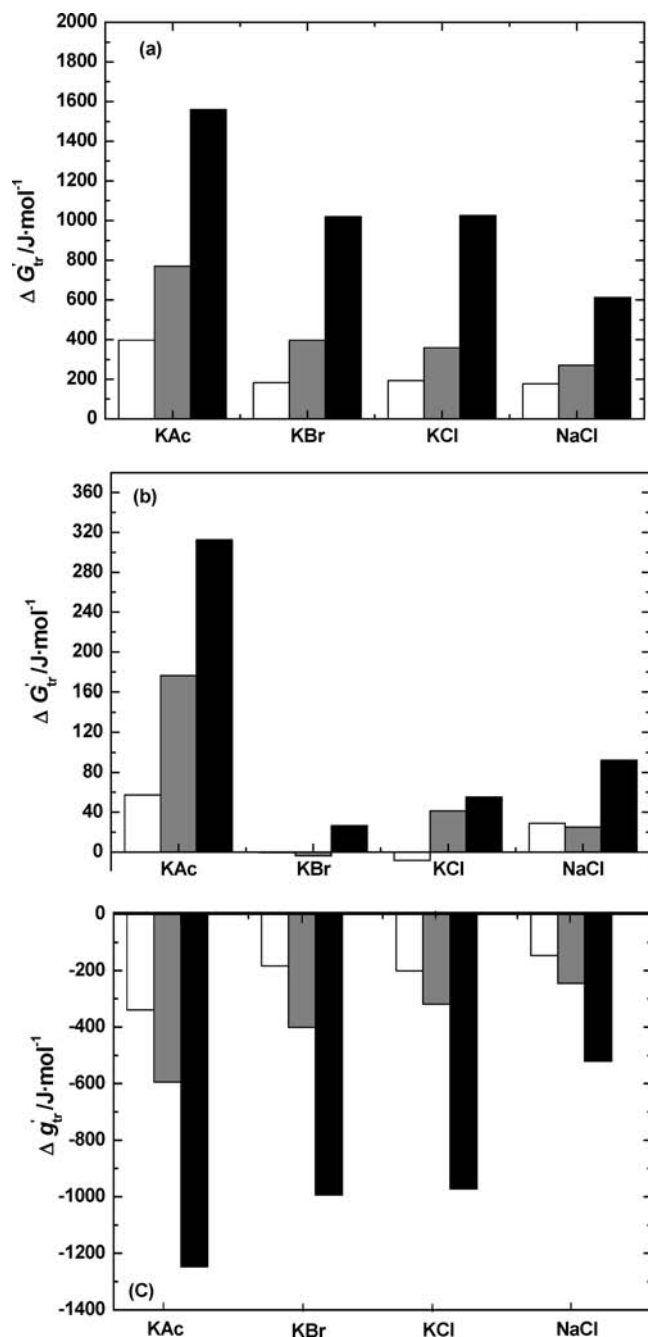
and  $\gamma_{B,w}$  and  $\gamma_{B,ws}$  are the activity coefficients of the solute expressed in terms of molar, molal, or mole fraction in water and in salt solution, respectively, at the solubility limit. The transfer free energy of the biological buffer from water to salt solution is equal to the difference between the standard state chemical potentials of the buffer in each solution, eq 8.<sup>37</sup>

$$\Delta\mu^o = \mu_{B,ws}^o - \mu_{B,w}^o = RT \ln \left( \frac{S_{B,w}}{S_{B,ws}} \right) + RT \ln \left( \frac{\gamma_{B,w}}{\gamma_{B,ws}} \right) \quad (8)$$

According to Nozaki and Tanford's analysis,<sup>32-34</sup> the ratio of the activity coefficient term makes only a small contribution to  $\Delta G_{tr}$ , thereby this effect of the simplification is negligible and also not much greater than the experimental uncertainty. Many researchers<sup>32-34,42,43</sup> have ignored the activity coefficient on the right-hand side of eq 8. We used only the first term on the right side of eq 8 and declare the quantities reported here as apparent transfer free energy ( $\Delta G'_{tr}$ ).<sup>32-34</sup> The apparent transfer free energy expressed in unit of molarity is shown in eq 9.<sup>37</sup>

$$\Delta G'_{tr} = RT \ln \left( \frac{n_{B,w}}{n_{B,ws}} \right) + RT \ln \left( \frac{V_{S,ws}}{V_{S,w}} \right) \quad (9)$$

Here,  $n_{B,w}$  and  $n_{B,ws}$  represent the moles of solute soluble in 100 g of water and in 100 g of electrolyte solution and  $V_{S,w}$  and  $V_{S,ws}$  are the total volumes of the aqueous solution and electrolyte solution containing the saturating solute on the molarity scale. It should be noted that the transfer free energies reported by Cohn and Edsall<sup>45</sup> and Tanford<sup>46</sup> are based upon the mole fraction scale, while Robinson and Jencks<sup>47,48</sup> and Bolen and co-workers<sup>49,50</sup> used the molarity scale. Ben-Naim<sup>51</sup> has strongly suggested in the favor of molarity scale for obtaining transfer free energies. Therefore, we also used molarity

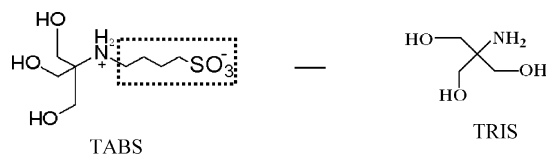


**Figure 2.** Display of apparent transfer free energies of (a) TRIS; (b) TABS; (c) TABS residue contribution from water to 1 M (white), 2 M (gray), and 4 M (black) of the salts at  $T = 298.15$  K.

scale for calculating the transfer free energies (eq 9). The calculated  $\Delta G'_{tr}$  values are reported in Table 5.

The transfer model has been a fixture in biophysical chemistry since at least the 1930s.<sup>52</sup> Usually, the transfer free energy is positive indicating that the interactions between the biological buffer and the salt are unfavorable. In contrast, if  $\Delta G'_{tr}$  has a negative sign, it indicates that the interactions are favorable. Table 5 and Figure 2 (a) illustrate the unfavorable interactions of TRIS buffer with four salts, positive values of  $\Delta G'_{tr}$ , and increases with increasing salt concentration. The positive  $\Delta G'_{tr}$  values indicate that salt has a salting-out effect on the hydrophobic group. Interestingly, a large increase in the  $\Delta G'_{tr}$  for TRIS occurred in KAc solution. It should also be noted that the interactions of TABS with salts are much less unfavorable than those on TRIS (Figure 2 (b)); the zwitterions have very

### Scheme 2. Schematic Illustration of the Contribution of the TABS Side Chain



large dipole moments<sup>53–55</sup> and interact electrostatically with the solvent and other charged molecules in the solution.<sup>29,56–60</sup> Surprisingly, the negative  $\Delta G'_{tr}$  values are obtained in TABS with (1 and 2)  $\text{mol}\cdot\text{dm}^{-3}$  KBr or 1  $\text{mol}\cdot\text{dm}^{-3}$  KCl, favorable interactions, and this behavior is sharply reversed since these solutions exhibit a salting-out effect.

The  $\Delta G'_{tr}$  values obtained from this study can be used to shed more light on the interaction between TABS and salts, by estimating the contribution of the side chain of TABS to the transfer free energy,  $\Delta g'_{tr}$ , assuming additivity of the free energy of solvent interactions. The side chain contributions to the transfer free energies of TABS are obtained simply from the difference between  $\Delta G'_{tr}$  of the TABS and those of TRIS at the same experimental conditions, as illustrated by Scheme 2.

Since  $\Delta g'_{tr}$  is, by definition, free of solute–solute interactions, it provides information regarding solute–solvent interactions. The results in Table 5 and graphically illustrated in Figure 2 (c) reveal that the apparent transfer energy ( $\Delta g'_{tr}$ ) contributions of TABS residue from water to aqueous electrolyte solutions (from Scheme 2) are negative and also increase with increasing salt concentration. The negative contribution indicates that the interactions between the solvent and TABS residue are favorable.

The apparent molar volumes,  $V_\phi$  of TRIS and TABS in water and in aqueous electrolyte solution were computed from the density of the solution by using the following equation

$$V_\phi = (M/\rho) - [10^3(\rho - \rho_0)/(m\rho\rho_0)] \quad (10)$$

where  $M$  and  $m$  are, respectively, the molar mass and molality of the biological buffers and  $\rho$  and  $\rho_0$  are the densities of solution and the solvent (water + ionic salts, or water), respectively. The values of  $V_\phi$  at each ionic salt concentration at (298.15 to 328.15) K are summarized in Tables 6 and 7. It was found that  $V_\phi$  values of TRIS and TABS increase with increasing temperature in the presence of all cosolutes studied. We fitted the apparent molar volumes  $V_\phi$  to empirical function (eq 11) of  $m$  and  $T$  for each system

$$V_\phi(m, T) = v_0 + v_1T + v_2T^2 + v_3mT^5 + v_4m + v_5m^2 + \frac{v_6m}{(m^5T - 273.15)} \quad (11)$$

In eq 11,  $v_i$  are regression coefficients given in Table 8 with average absolute deviation (AARD).

To shed more light on the interactions between the biological buffers under study and salts, we studied the partial molar volumes,  $V_\phi^0$ , of biological buffers in water at infinite dilution from the solubility data at 298.15 K, following the procedure of Auton et al.<sup>61</sup>

Ford et al.<sup>62</sup> measured apparent molar volumes of TRIS at molalities  $m \leq 0.5 \text{ mol}\cdot\text{kg}^{-1}$  and at temperatures from (278.15 to 393.15) K well below the minimum  $m = 1.6 \text{ mol}\cdot\text{kg}^{-1}$  of our experiments. Our  $V_\phi^0$  value ( $9.17 \text{ cm}^3\cdot\text{mol}^{-1}$ ) at  $T = 298.15$





**Table 7. Apparent Molar Volumes  $V_\phi$  of TABS in Water and in Aqueous Electrolyte Solutions as a Function of Concentration of TABS and Ionic Salts from (298.15 to 328.15) K under Atmospheric Pressure**

$m$ mol·kg <sup>-1</sup>	$V_\phi/\text{cm}^3\cdot\text{mol}^{-1}$						$m$ mol·kg <sup>-1</sup>	$V_\phi/\text{cm}^3\cdot\text{mol}^{-1}$					
	298.15 K	303.15 K	310.15 K	318.15 K	323.15 K	328.15 K		298.15 K	303.15 K	310.15 K	318.15 K	323.15 K	328.15 K
TABS in water							TABS in 1 mol·dm <sup>-3</sup> KAc						
0.9378	170.75	171.30	171.98	172.67	173.12	174.56	1.1092	168.89	169.30	169.86	170.43	170.72	170.82
1.5515	172.86	173.35	174.03	174.71	175.13	176.00	1.7143	174.20	174.61	175.20	175.79	176.10	176.22
2.4874	173.34	173.85	174.50	175.17	175.55	175.92	2.0187	175.37	175.83	176.45	177.09	177.43	177.61
4.0801	172.68	173.15	173.76	174.39	174.77	175.12	2.2756	174.25	174.70	175.31	175.94	176.28	176.49
4.7956	173.16	173.61	174.19	174.81	175.18	175.54	3.0564	176.74	177.19	177.79	178.43	178.78	179.03
5.1539	172.76	173.20	173.78	174.40	174.76	175.11	3.7151	175.03	175.46	176.03	176.64	176.99	177.24
5.5488	173.26	173.70	174.28	174.90	175.27	175.62	4.7178	175.15	175.55	176.10	176.68	177.02	177.29
							5.1065	174.74	175.14	175.69	176.27	176.61	176.88
							5.4501	174.24	174.63	175.16	175.74	176.07	176.33
TABS in 2 mol·dm <sup>-3</sup> KAc							TABS in 4 mol·dm <sup>-3</sup> KAc						
0.7450	179.75	180.19	180.77	181.33	181.65	181.99	0.6829	168.32	168.48	168.62	168.71	168.91	169.09
1.0801	176.79	177.21	177.80	178.42	178.76	179.07	1.0999	173.87	174.11	174.41	174.68	174.88	175.06
1.2927	177.27	177.76	178.39	179.04	179.39	179.71	1.4753	174.65	174.93	175.35	175.79	176.02	176.25
1.4913	177.64	178.13	178.78	179.45	179.81	180.14	1.9887	176.89	177.20	177.64	178.13	178.40	178.66
1.9129	173.96	174.42	175.02	175.65	175.98	176.30	2.8255	178.93	179.26	179.72	180.23	180.53	180.82
2.7415	175.20	175.62	176.17	176.77	177.10	177.42	3.2752	179.26	179.58	180.04	180.56	180.87	181.16
3.3739	174.49	174.88	175.42	176.00	176.33	176.64	3.9265	178.63	178.95	179.40	179.93	180.24	180.54
3.8714	174.83	175.21	175.74	176.31	176.64	176.95							
4.4438	174.91	175.28	175.80	176.35	176.68	176.99							
TABS in 1 mol·dm <sup>-3</sup> KBr							TABS in 2 mol·dm <sup>-3</sup> KBr						
1.1889	171.85	172.28	172.83	173.38	173.42	173.52	0.9366	173.45	173.82	174.34	174.90	175.07	175.20
1.7909	173.41	173.86	174.38	174.94	175.07	175.28	1.6871	175.22	175.59	176.09	176.65	176.90	177.23
2.3552	172.63	173.05	173.60	174.16	173.38	173.33	2.2775	174.07	174.44	174.94	175.50	175.77	175.85
3.0462	175.22	175.63	176.18	176.75	176.24	176.31	3.3510	174.41	174.78	175.28	175.83	176.13	176.23
3.7081	173.30	173.70	174.24	174.80	174.90	175.04	4.3871	174.68	175.04	175.54	176.10	176.40	176.48
4.4738	173.99	174.39	174.91	175.47	175.54	175.62	4.7318	174.12	174.47	174.97	175.51	175.81	175.91
TABS in 4 mol·dm <sup>-3</sup> KBr							TABS in 1 mol·dm <sup>-3</sup> KCl						
0.9627	176.49	176.78	177.22	177.72	178.03	178.32	1.2651	171.94	172.40	172.97	173.59	173.96	174.30
1.4267	175.94	176.25	176.69	177.19	177.50	177.80	1.8927	173.47	173.91	174.49	175.10	175.46	175.80
2.0066	175.34	175.65	176.10	176.60	176.92	177.23	2.5873	173.43	173.87	174.44	175.06	175.42	175.77
3.0140	175.47	175.80	176.25	176.76	177.08	177.39	3.6148	175.18	175.59	176.13	176.72	177.08	177.44
3.6440	175.62	175.95	176.40	176.91	177.23	177.54	4.2165	173.10	173.51	174.05	174.64	175.00	175.35
3.9499	175.41	175.73	176.17	176.68	177.00	177.31	4.8710	173.66	174.07	174.60	175.19	175.55	175.90
							5.1605	173.02	173.43	173.97	174.55	174.91	175.25
TABS in 2 mol·dm <sup>-3</sup> KCl							TABS in 4 mol·dm <sup>-3</sup> KCl						
1.0008	167.25	167.65	168.17	168.70	169.05	169.42	0.9914	172.19	172.52	172.70	173.06	173.38	173.70
1.6902	172.74	173.13	173.65	174.23	174.60	174.95	1.5752	174.03	174.36	174.80	175.26	175.59	175.93
2.4232	171.86	172.25	172.77	173.33	173.68	174.02	2.2794	175.09	175.44	175.93	176.48	176.82	177.16
3.1228	174.41	174.80	175.33	175.91	176.26	176.61	2.9153	175.88	176.23	176.73	177.29	177.63	177.97
3.8344	172.89	173.27	173.78	174.35	174.69	175.02	3.5939	174.80	175.14	175.63	176.18	176.52	176.86
4.5115	173.09	173.47	173.97	174.53	174.87	175.21	4.0249	175.50	175.85	176.33	176.89	177.23	177.57
4.8158	172.68	173.05	173.56	174.12	174.46	174.79	4.3125	174.93	175.27	175.75	176.31	176.65	176.99
TABS in 1 mol·dm <sup>-3</sup> NaCl							TRIS in 2 mol·dm <sup>-3</sup> NaCl						
0.9211	172.65	173.11	173.70	174.32	174.65	174.78	1.0194	173.59	173.97	174.51	175.08	175.44	175.78
1.6553	175.57	175.97	176.53	177.10	177.43	177.14	1.7625	174.80	175.16	175.69	176.26	176.62	176.96
2.0066	175.61	176.02	176.59	177.19	177.54	177.64	2.1442	174.01	174.38	174.91	175.47	175.84	176.19
2.4096	174.38	174.81	175.39	176.01	176.36	176.46	2.7734	176.03	176.41	176.94	177.51	177.88	178.22
3.8585	173.72	174.13	174.68	175.28	175.63	175.70	3.6141	174.37	174.75	175.26	175.82	176.17	176.51
4.6879	174.59	175.00	175.53	176.13	176.48	176.60	4.4423	175.16	175.53	176.03	176.60	176.94	177.28
5.3560	174.13	174.53	175.06	175.65	176.00	176.15	4.7936	174.61	174.98	175.48	176.04	176.39	176.73
TRIS in 4 mol·dm <sup>-3</sup> NaCl													
0.9452	175.77	176.09	176.54	177.10	177.43	177.75							
1.5200	176.72	177.06	177.52	178.08	178.41	178.74							
1.8185	175.56	175.90	176.37	176.92	177.25	177.58							
2.2643	176.93	177.28	177.76	178.32	178.65	179.00							
3.1003	175.64	175.98	176.46	177.00	177.33	177.66							
3.9740	175.63	175.96	176.44	176.98	177.31	177.65							
4.3066	175.59	175.93	176.39	176.93	177.27	177.60							

KCl with increasing concentration, ended by a negative value at 4 mol·dm<sup>-3</sup> KCl. The  $\Delta_{\text{tr}}V_\phi^\circ$  (H<sub>2</sub>O aqueous KAc and NaCl) of TRIS is negative at first, then positive with an increase of molarity of salt. The positive  $\Delta_{\text{tr}}V_\phi^\circ$  values for TRIS and TABS indicate that ion–hydrophilic interactions predominate over the ion–hydrophobic interactions. Interestingly, the transfer volumes  $\Delta_{\text{tr}}V_\phi^\circ$  of TRIS are less than 50 % of that of

TABS. It may be due to the fact that the interaction between zwitterionic end groups of TABS and ions in solvent becomes stronger.

Assuming that there is no interaction between the groups of biological buffers, the contributions of groups to the  $\Delta_{\text{tr}}V_\phi^\circ$  are additive, as we have mentioned before. The  $\Delta_{\text{tr}}V_\phi^\circ$  ( $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{SO}_3^-$ ) approaches  $\Delta_{\text{tr}}V_\phi^\circ(\text{TABS}) - \Delta_{\text{tr}}V_\phi^\circ$

**Table 8. Regression Parameters of Equation 9 for Apparent Molar Volumes  $V_\phi^0$  and Also the Apparent Molar Volume at Infinities Dilution  $V_\phi^0$  at 298.15 K of TRIS and TABS in Water and in Aqueous Electrolyte Solutions**

system	$v_0$	$10^2 \cdot v_1$	$10^5 \cdot v_2$	$10^{14} \cdot v_3$	$v_4$	$v_5$	$v_6$	AARD/% <sup>a</sup>
water + TRIS	74.80	0.05699	-0.32	1.6	0.5524	-0.107	-6940	0.13
water + TRIS + 1 M KAc	88.40	-0.7710	6.32	12.4	1.2571	-0.409	-1	0.24
water + TRIS + 2 M KAc	39.12	25.2220	-30.60	-1.7	6.2647	-1.462	17	0.94
water + TRIS + 4 M KAc	156.25	-52.6147	93.81	-15.3	15.0753	-5.042	-197	1.44
water + TRIS + 1 M KBr	-808.81	590.9129	-985.29	185.5	9.5963	-2.757	-145	2.10
water + TRIS + 2 M KBr	-507.90	390.1630	-641.86	113.4	4.8421	-1.649	-401	1.22
water + TRIS + 4 M KBr	125.66	-3.6640	-10.21	102.2	-11.3883	1.494	-30	0.05
water + TRIS + 1 M KCl	306.44	-141.2780	232.38	1.7	3.1613	-0.667	2	0.91
water + TRIS + 2 M KCl	47.86	16.9087	-25.69	23.7	12.5485	-2.278	-427	0.12
water + TRIS + 4 M KCl	195.60	-43.9027	64.20	89.2	-39.1558	10.096	299	2.22
water + TRIS + 1 M NaCl	-56.91	88.0763	-139.33	21.7	7.9488	-1.435	-4	1.67
water + TRIS + 2 M NaCl	50.66	17.2222	-24.24	18.0	9.2384	-1.649	-251	1.04
water + TRIS + 4 M NaCl	187.38	-47.5780	50.83	145.0	0.8926	-1.197	-163	1.23
water + TABS	177.40	-14.9990	44.85	-22.6	0.6081	0.007	94	0.15
water + TABS + 1 M KAc	73.24	57.8460	-83.52	8.5	2.4606	-0.401	-1119	0.24
water + TABS + 2 M KAc	115.44	35.5890	-44.32	-3.5	-4.5085	0.677	-176	0.29
water + TABS + 4 M KAc	125.01	21.9910	-31.35	26.6	9.7276	-1.618	372	0.07
water + TABS + 1 M KBr	-246.26	270.0360	-437.00	43.6	1.1603	-0.339	-267	0.38
water + TABS + 2 M KBr	-94.94	169.6950	-264.00	7.1	-1.3034	0.151	104	0.19
water + TABS + 4 M KBr	146.18	14.9230	-14.72	4.1	-1.6028	0.251	-0.5	0.08
water + TABS + 1 M KCl	120.30	24.6390	-26.85	-1.6	2.6008	-0.379	-316	0.23
water + TABS + 2 M KCl	153.95	8.6404	-11.56	30.3	1.9194	-0.414	-138	0.34
water + TABS + 4 M KCl	157.62	3.8888	0.13	16.9	3.68187	-0.653	-10	0.16
water + TABS + 1 M NaCl	9.76	100.7967	-148.86	-3.5	-1.9076	0.245	272	0.18
water + TABS + 2 M NaCl	138.89	14.0916	-10.52	-2.1	2.3028	-0.347	6	0.27
water + TABS + 4 M NaCl	163.13	1.9213	9.37	-5.8	-0.5232	0.073	36	0.18

<sup>a</sup> AARD/% =  $(100/N) \sum_{i=1}^N |V_{\phi,i}^{\text{calc}} - V_{\phi,i}^{\text{expt}}| / V_{\phi,i}^{\text{expt}}$ , where  $N$  is the number of data points.

**Table 9. Apparent Molar Volume at Infinite Dilution  $V_\phi^0$  and Partial Molar Volumes  $\Delta_{\text{tr}}V_\phi^0$  for TRIS and TABS in Aqueous Electrolyte Solutions at  $T = 298.15$  K and at Atmospheric Pressure**

solvent	$V_\phi^0/\text{cm}^3 \cdot \text{mol}^{-1}$				$\Delta_{\text{tr}}V_\phi^0/\text{cm}^3 \cdot \text{mol}^{-1}$		
	0 mol·dm <sup>-3</sup>	1 mol·dm <sup>-3</sup>	2 mol·dm <sup>-3</sup>	4 mol·dm <sup>-3</sup>	1 mol·dm <sup>-3</sup>	2 mol·dm <sup>-3</sup>	4 mol·dm <sup>-3</sup>
TRIS							
water	91.17						
KAc		91.14	91.89	92.17	-0.03	0.72	1.00
KBr		91.34	91.63	92.57	0.17	0.46	1.40
KCl		91.85	91.31	88.15	0.68	0.14	-3.02
NaCl		90.95	91.94	92.23	-0.22	0.77	1.06
TABS							
water	169.24						
KAc		172.95	171.10	177.5	3.71	1.86	8.26
KBr		171.76	173.19	174.78	2.52	3.95	5.54
KCl		171.07	171.12	173.65	1.83	1.88	4.41
NaCl		171.14	172.46	173.94	1.90	3.22	4.70

**Table 10. Contribution of TABS Residue ( $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{SO}_3^-$ ) to the Partial Molar Volumes of Transfer,  $\Delta_{\text{tr}}V_\phi^0$ , in Aqueous Electrolyte at  $T = 298.15$  K and at Atmospheric Pressure**

system	$\Delta_{\text{tr}}V_\phi^0/\text{cm}^3 \cdot \text{mol}^{-1}$		
	1 mol·dm <sup>-3</sup>	2 mol·dm <sup>-3</sup>	4 mol·dm <sup>-3</sup>
KAc	3.74	1.14	7.26
KBr	2.35	3.49	4.14
KCl	1.15	1.74	7.43
NaCl	2.12	2.45	3.64

(TRIS) (Scheme 2). Table 10 shows the contribution of TABS residue is positive, which indicates that the ion-hydrophilic interaction between the ions salt and sulfonic group is predominate over the ion-hydrophobic interactions between the ion salt and methyl groups of TABS residue.

## Conclusions

This work provides reliable density data for various binary and ternary aqueous systems that contain TRIS or TABS and electrolytes such as KAc, KBr, KCl, and NaCl from (298.15 to 328.15) K under atmospheric pressure. The experimental data were correlated satisfactorily with an empirical equation

in terms of the concentrations of buffer and salts, with an augmented term to account for the interactions between buffer and electrolytes. The solubility limits of TRIS and TABS in the aqueous or the electrolyte solutions were determined at 298.15 K from the results of the density measurement. The solubilities of TABS in aqueous solution decrease with increasing concentration of salt (salting-out effect). The observed order of the salting-out effect is KAc > KBr > KCl > NaCl for TRIS and KAc > KBr > KCl  $\approx$  NaCl for TABS, respectively. We have determined the apparent transfer free energies ( $\Delta G_{\text{tr}}^0$ ) of the biological buffers from water to aqueous electrolyte solutions based on the solubility data. The values of  $\Delta G_{\text{tr}}^0$  were used also to calculate the contributions of TABS residue ( $-\text{CH}_2\text{CH}_2\text{CH}_2\text{CH}_2\text{SO}_3^-$ ) from water to aqueous electrolyte solutions. We have calculated the apparent molar volumes,  $V_\phi(m, T)$ , from the solubility data. The  $\Delta_{\text{tr}}V_\phi^0$  values are almost positive indicating the dominance of ion-hydrophilic interactions. Also, TABS residue contributions calculated from  $V_\phi^0$  suggest that ion-hydrophilic interaction between the ion salts and sulfonic group overwhelms the hydrophobic interactions between the ion salt and methyl groups of TABS residue.

Overall, these results provide evidence for the existence of interactions between the buffers and ionic salts, in aqueous solutions. It was found that the salt effects on TABS are greater than those on TRIS.

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### Literature Cited

- Thiela, T.; Liczkowski, L.; Bissena, S. T. New Zwitterionic Butane-sulfonic Acids that Extend the Alkaline Range of Four Families of Good Buffers: Evaluation for Use in Biological Systems. *J. Biochem. Biophys. Methods* **1998**, *37*, 117–129.
- Halford, S. E.; Goodall, A. J. Modes of DNA Cleavage by the EcoRV Restriction Endonuclease. *Biochemistry* **1988**, *27*, 1771–1777.
- Good, N. E.; Winget, G. D.; Winter, W.; Connolly, T. N.; Izawa, S.; Singh, R. M. M. Hydrogen Ion Buffers for Biological Research. *Biochemistry* **1966**, *5*, 467–477.
- Ferguson, W. J.; Braunschweiger, K. I.; Braunschweiger, W. R.; Smith, J. R.; McCormick, J. J.; Wasmann, C. C.; Jarvis, N. P.; Bell, D. H.; Good, N. E. Hydrogen Ion Buffers for Biological Research. *Anal. Biochem.* **1980**, *104*, 300–310.
- Gomori, G. Preparation of Buffers for Use in Enzyme Studies. *Methods Enzymol.* **1955**, *1*, 138–146.
- Brzozowski, A. M.; Lawson, D. M.; Turkenburg, J. P.; Bisgaard-Frantzen, H.; Svendsen, A.; Borchert, T. V.; Dauter, Z.; Wilson, K. S.; Davies, G. J. Structural Analysis of a Chimeric Bacterial  $\alpha$ -Amylase. High-Resolution Analysis of Native and Ligand Complexes. *Biochemistry* **2000**, *39*, 9099–9107.
- Knapp, S.; Rüdiger, A.; Antranikian, G.; Ladenstein, R. Crystallization and Preliminary Crystallographic Analysis of an Amyloplullulanase from the Hyperthermophilic Archaeon *Pyrococcus Woesei*. *Proteins* **1995**, *23*, 595–597.
- Andrykovitch, M.; Guo, W.; Karen, M. R.; Gu, Y.; Anderson, D. E.; Reshetnikova, L. S.; Knowlton, J. R.; Waugh, D. S.; Ji, X. Crystallization and Preliminary x-Ray Diffraction Studies of NusG, a Protein Shared by the Transcription and Translation Machines. *Acta Crystallogr. D Biol. Crystallogr.* **2002**, *58*, 2157–2158.
- Campos, A.; Matsumura, P.; Volz, K. Crystallization and Preliminary x-Ray Analysis of FlhD from *Escherichia Coli*. *J. Struct. Biol.* **1998**, *123*, 269–271.
- Iguer-Ouada, M.; Versteegen, J. P. Long Term Preservation of Chilled Canine Semen: Effect of Commercial and Laboratory Prepared Extenders. *Theriogenology* **2000**, *55*, 671–84.
- Tsutsui, T.; Hase, M.; Hori, T.; Ito, T.; Kawakami, E. Effects of Orvus ES Paste on Canine Spermatzoal Longevity after Freezing and Thawing. *J. Vet. Med. Sci.* **2000**, *62*, 533–535.
- Silva, A. R.; Cardoso, R. C. S.; Uchoa, D. C.; Silva, L. D. M. Effect of Tris-buffer, Egg Yolk and Glycerol on Canine Semen Freezing. *Vet. J.* **2002**, *164*, 244–246.
- Lesignoli, F.; Germini, A.; Corradini, R.; Sforza, S.; Galaverna, G.; Dossena, A.; Marchelli, R. Recognition and Strand Displacement of DNA Oligonucleotides by Peptide Nucleic Acids (PNAs). High-performance Ion Exchange Chromatographic Analysis. *J. Chromatogr. A* **2001**, *922*, 177–185.
- Smith, W. H., Jr.; Hood, D. W. *pH Measurement in the Ocean: A Sea Water Secondary Buffer System. In Recent Researches in the Fields of Hydrosphere, Atmosphere and Nuclear Geochemistry*; Miyake, Y., Koyama, T., Eds.; Maruzen Company, Ltd: Tokyo, Japan, 1964; pp 185–202.
- Hansson, I. A New Set of pH Scales and Standard Buffers for Sea Water. *Deep-Sea Res.* **1973**, *20*, 479–491.
- Ramette, R. W.; Culbertson, C. H.; Bates, R. G. Acid-Base Properties of tris(Hydroxymethyl)aminomethane (Tris) Buffers in Seawater from 5 to 40°C. *Anal. Chem.* **1977**, *49*, 867–870.
- Dickson, A. G. pH Buffers for Sea Water Media Based on the Total Hydrogen Ion Concentration Scale. *Deep-Sea Res.* **1993**, *40*, 107–118.
- DelValls, T. A.; Dickson, A. G. The pH of Buffers Based on 2-Amino-2-hydroxymethyl-1,3-propanediol (Tris) in Synthetic Sea Water. *Deep-Sea Res.* **1998**, *45*, 1541–1554.
- Whitfield, M.; Butler, R. A.; Covington, A. K. The Determination of pH in Estuarine Waters: I. Definition of pH Scales and the Selection of Buffers. *Oceanologica Acta* **1985**, *8*, 423–432.
- Millero, F. J.; Zhang, J.-Z.; Fiol, S.; Sotolongo, S.; Roy, R. N.; Lee, K.; Mane, S. The Use of Buffers to Measure the pH of Seawater. *Mar. Chem.* **1993**, *44*, 143–152.
- Clayton, T. D.; Byrne, R. H. Spectrophotometric Seawater pH Measurements: Total Hydrogen Ion Concentration Scale Calibration of m-Cresol Purple and at-Sea Results. *Deep-Sea Res.* **1993**, *40*, 2115–2129.
- Zhang, H.; Byrne, R. H. Spectrophotometric pH Measurements of Surface Seawater at In-Situ Conditions: Absorbance and Protonation Behavior of Thymol Blue. *Mar. Chem.* **1996**, *52*, 17–25.
- Nakaguchi, Y.; Nemzer, B. V.; Dickson, A. G. The Calibration of Indicator Dyes for Seawater pH Measurements. In *6th International Carbon Dioxide Conference*, Sendai, Japan, 2001; pp 631–634.
- Stellwagen, N. C.; Gelfi, C.; Righetti, P. G. The Use of Gel and Capillary Electrophoresis to Investigate Some of the Fundamental Physical Properties of DNA. *Electrophoresis* **2002**, *23*, 167–175.
- Stellwagen, N. C.; Gelfi, C.; Righetti, P. G. The Free Solution Mobility of DNA. *Biopolymers* **1997**, *42*, 687–703.
- Stellwagen, N. C.; Bossi, A.; Gelfi, C.; Righetti, P. G. DNA and Buffers: Are There Any Noninteracting, Neutral pH Buffers. *Anal. Biochem.* **2000**, *287*, 167–175.
- Stellwagen, E.; Stellwagen, N. C. The Free Solution Mobility of DNA in Tris-acetate-EDTA Buffers of Different Concentrations, with and without Added NaCl. *Electrophoresis* **2002**, *23*, 1935–1941.
- Stellwagen, N. C.; Gelfi, C.; Righetti, P. G. DNA-Histidine Complex Formation in Isoelectric Histidine Buffers. *J. Chromatogr. A* **1999**, *838*, 179–189.
- Wenner, J. R.; Bloomfield, V. A. Buffer Effects on EcoRV Kinetics as Measured by Fluorescent Staining and Digital Imaging of Plasmid Cleavage. *Anal. Biochem.* **1999**, *268*, 201–212.
- Ashcroft, S. J.; Booker, D. R.; Robin Turner, J. C. Density Measurement by Oscillating Tube. Effects of Viscosity, Temperature, Calibration and Signal Processing. *J. Chem. Soc., Faraday Trans.* **1990**, *86*, 145–149.
- Fandino, O.; Garcia, J.; Comunas, M. J. P.; Lopez, E. R.; Fernandez, J. P. T.  $\rho$  T Measurements and Equation of State (EoS) Predictions of Ester Lubricants up to 45 MPa. *Ind. Eng. Chem. Res.* **2006**, *45*, 1172–1182.
- Nozaki, Y.; Tanford, C. The Solubility of Amino Acids and Related Compounds in Aqueous Urea Solutions. *J. Biol. Chem.* **1963**, *238*, 4074–4081.
- Nozaki, Y.; Tanford, C. The Solubility of Amino Acids and Related Compounds in Aqueous Ethylene Glycol Solutions. *J. Biol. Chem.* **1965**, *240*, 3568–3573.
- Nozaki, Y.; Tanford, C. The Solubility of Amino Acids, Diglycine, and Triglycine in Aqueous Guanidine Hydrochloride Solutions. *J. Biol. Chem.* **1970**, *245*, 1648–1652.
- Venkatesu, P.; Lee, M. J.; Lin, H. M. Thermodynamic Characterization of the Osmolyte Effect on Protein Stability and the Effect of GdnHCl on Protein Denatured State. *J. Phys. Chem. B* **2007**, *111*, 9045–9056.
- Venkatesu, P.; Lee, M. J.; Lin, H. M. Trimethylamine *N*-oxide Counteracts the Denaturing Effects of Urea or GdnHCl on Protein Denatured State. *Arch. Biochem. Biophys.* **2007**, *466*, 106–115.
- Auton, M.; Bolen, D. W. Additive Transfer Free Energies of the Peptide Backbone Unit that Are Independent of the Model Compound and the Choice of Concentration Scale. *Biochemistry* **2004**, *43*, 1329–1342.
- Schindler, P.; Robinson, R. A.; Bates, R. G. Solubility of tris(Hydroxymethyl)aminomethane in Water-methanol Solvent Mixtures and Medium Effects in Dissociation of Protonated Base. *J. Res. Natl. Bur. Stand.* **1968**, *72A*, 141–148.
- El-Harakany, A. A.; Barakat, A. O. Solubility of tris-(Hydroxymethyl)-aminomethane in Water-2-methoxyethanol Solvent Mixtures and the Solvent Effect on the Dissociation of the Protonated Base. *J. Solution Chem.* **1985**, *14*, 263–269.
- Taha, M.; Lee, M. J. Interaction of Biological Buffers with Electrolytes: Densities of Aqueous Solutions of two Substituted Aminosulfonic Acids and Ionic Salts from  $T = (298.15 \text{ to } 328.15) \text{ K}$ . *J. Chem. Thermodyn.* **2009**, *41*, 705–715.
- Edsall J. T.; Wyman, J. *Biophysical Chemistry*; Academic Press: London, 1958, Vol. I.
- Murphy, K. P. *Protein structure, Stability, and Folding*; Humana Press: Totowa, NJ, 2001.
- Lapaanje, S.; Skerjance, J.; Glavnik, S.; Zibret, S. Thermodynamic Studies of the Interactions of Guanidinium Chloride and Urea with Some Oligoglycines and Oligoalucines. *J. Chem. Thermodyn.* **1978**, *10*, 425–433.
- Pittz, E. P.; Bello, J. Studies on Bovine Pancreatic Ribonuclease A and Model Compounds in Aqueous 2-Methyl-2,4-pentandiol: I. Amino Acid Solubility, Thermal Reversibility of Ribonuclease A, and Preferential Hydration of Ribonuclease A Crystals. *Arch. Biochem. Biophys.* **1971**, *146*, 513–524.
- Cohn, E. J.; Edsall, J. T. *Proteins, Amino Acids, and Peptides as Ions and Dipolar Ions*; Reinhold Publishing Corporation: New York, 1943.

- (46) Tanford, C. *The Hydrophobic Effect: Formation of Micelles and Biological Membranes*; John Wiley and Sons: New York, 1973.
- (47) Robinson, D. R.; Jencks, W. P. Effect of Denaturing Agents of the Urea-Guanidinium Class on the Solubility of Acetyltetraglycine Ethyl Ester and Related Compounds. *J. Biol. Chem.* **1963**, *238*, 1558–1560.
- (48) Robinson, D. R.; Jencks, W. P. The Effect of Compounds of the Urea-Guanidinium Class on the Activity Coefficient of Acetyltetraglycine Ethyl Ester and related Compounds. *J. Am. Chem. Soc.* **1965**, *87*, 2462–2470.
- (49) Liu, Y.; Bolen, D. W. The Peptide Backbone Plays a Dominant Role in Protein Stabilization by Naturally Occurring Osmolytes. *Biochemistry* **1995**, *34*, 12884–12891.
- (50) Wang, A.; Bolen, D. W. Naturally Occurring Protective System in Urea-rich Cells: Mechanism of Osmolyte Protection of Proteins against Urea Denaturation. *Biochemistry* **1997**, *36*, 9101–9108.
- (51) Ben-Naim, A. Standard Thermodynamics of Transfer. Uses and Misuses. *J. Phys. Chem.* **1978**, *82*, 792–803.
- (52) McMeekin, T. L.; Cohn, E. J.; Wear, J. H. Studies in the Physical Chemistry of Amino Acids, Peptides and Related Substances. VII. A Comparison of the Solubility of Amino Acids, Peptides and Their Derivatives. *J. Am. Chem. Soc.* **1936**, *58*, 2173–2181.
- (53) Kirkwood, J. G. Theory of Solutions of Molecules Containing Widely Separated Charges with Special Application to Zwitterions. *J. Chem. Phys.* **1934**, *2*, 351–361.
- (54) Neuberger, A. Dissociation Constants and Structures of Zwitterions. *Proc. R. Soc. London A.* **1937**, *158*, 68–96.
- (55) Robinson, R. A.; Stokes, R. H. *Electrolyte Solutions*, 2nd ed.; Dover: Mineola, NY, 2002.
- (56) Roy, R. N.; Robinson, R. A.; Bates, R. G. Thermodynamics of the Two Dissociation Steps of *N*-tris(Hydroxymethyl)methylglycine (“Tricine”) in Water from 5 to 50°C. *J. Am. Chem. Soc.* **1973**, *95*, 8231–8235.
- (57) Cecchi, T.; Cecchi, P. The Dipole Approach in the Ion-Interaction Chromatography of Zwitterions: Use of a Potential Approximation to Obtain a Simplified Retention Equation. *Chromatographia* **2002**, *55*, 279–282.
- (58) Cecchi, T.; Pucciarelli, F.; Passamonti, P. Ion-Interaction Chromatography of Zwitterions. The Fractional Charge Approach to Model the Influence of the Mobile Phase Concentration of the Ion-Interaction Reagent. *Analyst* **2004**, *129*, 1037–1046.
- (59) Soto, A.; Arce, A.; Khoshkbarchi, M. K. Experimental Data and Modeling of Apparent Molar Volumes, Isentropic Compressibilities and Refractive Indices in Aqueous Solutions of Glycine + NaCl. *Biophys. Chem.* **1998**, *74*, 165–173.
- (60) Marcus, Y. On the Activity Coefficients of Charge-symmetrical Ion Pairs. *J. Mol. Liq.* **2006**, *123*, 8–13.
- (61) Auton, M.; Bolen, D. W.; Rösger, J. Structural Thermodynamics of Protein Preferential Salvation: Osmolyte Salvation of Proteins, Amino Acids, and Peptides. *Proteins: Struct., Funct., Bioinf.* **2008**, *73*, 802–812.
- (62) Ford, T. D.; Call, T. G.; Origlia, M. L.; Stark, M. A.; Woolley, E. M. Apparent Molar Volumes and Apparent Molar Heat Capacities of Aqueous 2-Amino-2-hydroxymethyl-propan-1,3-diol (Tris or THAM) and THAM Plus Equimolar HCl. *J. Chem. Thermodyn.* **2000**, *32*, 499–519.
- (63) Belibagli, K.; Agranci, E. Viscosities and Apparent Molar Volumes of Some Amino Acids in Water and in 6 M Guanidine Hydrochloride at 25°C. *J. Solution Chem.* **1990**, *19*, 867–882.
- (64) Friedman, H. L.; Krishnan, C. V. *Water: A Comprehensive Treatise*; Franks, F., Ed.; Plenum: New York, 1990; Vol. 35, pp 87–93.

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